

The Synthesis of a Terminally Linked Homodimeric Bisdistamycin Analog

Naim H. Al-Said*

Department of Applied Chemical Sciences, Jordan University of Science and Technology,
Irbid, Jordan

Received May 9, 2006; accepted May 24, 2006

Published online November 6, 2006 © Springer-Verlag 2006

Summary. A practical synthesis route to a terminally linked homodimeric bisdistamycin analog is described. In this analog the two strands of tricarboxamides of the pyrrole-pyrrole-pyrrole array are tethered from the nitrogen atom of the terminal pyrrole by a bisethoxyethane chain.

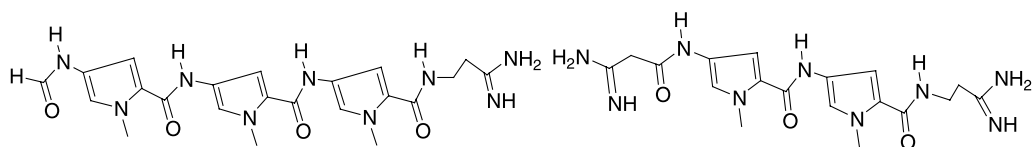
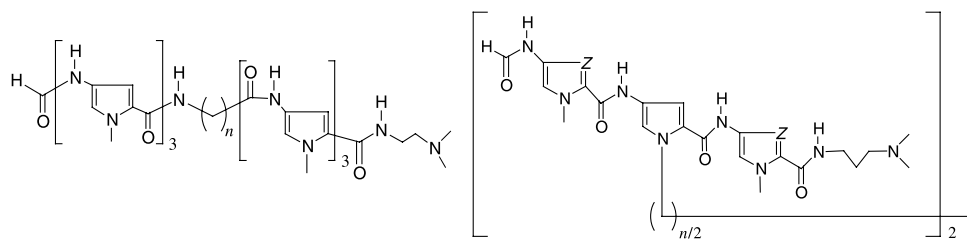
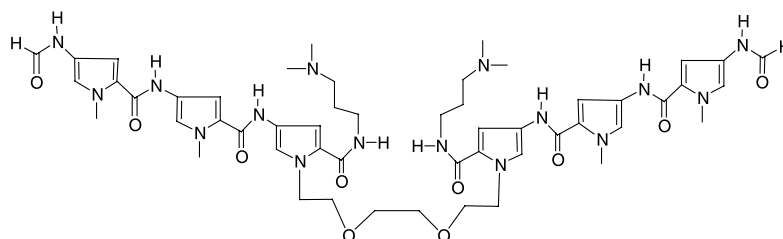
Keywords. Natural product; Stacking; *DNA*; Heterocyclic.

Introduction

The naturally occurring oligopeptide antibiotics distamycin (**1**) [1, 2] and netropsin (**2**) (Formulae 1) [3, 4] bind reversibly to specific nucleotide sequences consisting of 4–5 adjacent *AT* base pairs in the minor groove of double-helical *DNA* blocking its template function [5, 6]. Structural modifications by replacement of pyrrole by other heterocycles resulted in designing a novel class of minor groove binding agents called lexitropsins, *i.e.* “information reading molecules” [7–10]. The polypyrrolo-carboxamides skeletons of these oligopeptides have been used as *DNA* sequence selective vehicles for the delivery of alkylating agents to *DNA* targets [11–15].

Increasing the number of heterocyclic carboxamide units beyond five has resulted in compounds with lower binding affinity to *DNA*. This is due to the incompatible phasing of the crescent-shaped curvature, which prevents maximum hydrogen bonding and *Van der Waals* forces between the ligand and *DNA* [16–18]. Moreover, NMR studies have confirmed that two lexitropsin residues can be accommodated, stacked side-by-side in an antiparallel fashion, filling the minor groove of the *DNA* [19–22]. As a consequence, two types of bislexitropsin structures have been designed: a hairpin **3**, in which the two units of oligopeptides are covalently linked in head-to-tail fashion by a linker [23–25], and cross-linked **4**, where the central rings of the two oligopeptides are linked *via* a polymethylene chain (Formulae 2) [26–30]. These bislexitropsins recognize longer *DNA* sequences with

* E-mail: naim@just.edu.jo

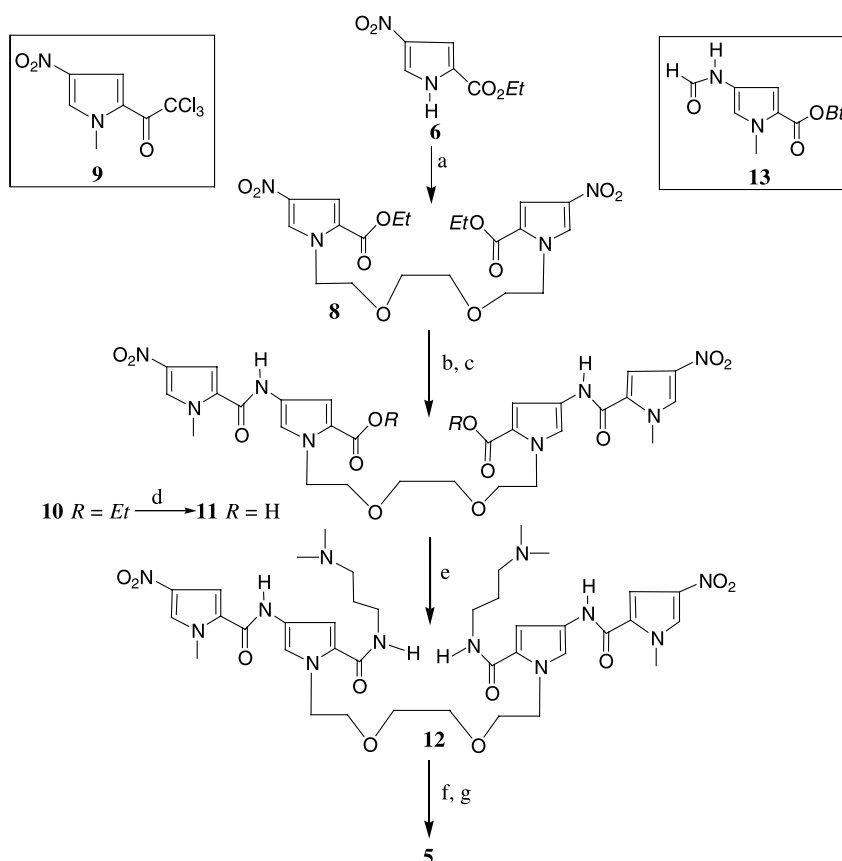
**1****2****Formulae 1****3****4** ($Z = \text{N}$ or CH ; $n = 5-10$)**5****Formulae 2**

stronger binding affinity and higher specificity compared with the monomer when the linker has the appropriate length. Additionally, these molecules also exhibit activity against retroviruses including HIV-1 [23–32]. Previously, we reported the synthesis of homodimeric cross-linked bis-lexitropsins **4** connected through the nitrogens of the two central pyrrole rings with polymethylene linkers [26, 27].

Herein, the synthesis of a third generation of bisdistamycin analog **5** is described. In this generation the two pyrrolo-carboxamide chains are connected through the nitrogen atoms of the two terminal pyrrole rings by a tether of a bis-ethoxyethane chain. The oxygen atoms are introduced in the chain to increase the solubility of the dimer in aqueous solution.

Results and Discussion

The key to the synthesis of terminally linked homodimeric bislexitropsins was the coupling of easily accessible ethyl 4-nitropyrrol-2-carboxylate (**6**) [31] and the commercially available bis(2-chloroethoxy)ethane (**7**) to form the central homo-



a) $\text{Cl}(\text{CH}_2)_2\text{O}(\text{CH}_2)_2\text{O}(\text{CH}_2)_2\text{Cl}$, LiI, *DMF*, 80°C , 2 h, 62%; b) H_2 , 5% Pd/C, 18 h, *MeOH*, *EtOAc*; c) **9**, *DMF*, Et_3N , 50°C , 3 h, 77% (2 steps); d) 2 *N* NaOH, *MeOH*, 50°C , 4 h, 71%; e) *EDCI*, *DMF*, $\text{H}_2\text{N}(\text{CH}_2)_3\text{NMe}_2$, Et_3N , *HOBT*, rt, 3 h; f) H_2 , 5% Pd/C, *MeOH*, rt, 6 h; g) **13**, Et_3N , *DMF*, 50°C , 68%

Scheme 1

unit of the 1,8-dipyrrrole derivative **8**. Initially, this compound was prepared in low yield by $\text{S}_{\text{N}}2$ reaction between **6** and **7** in *DMF* at ambient temperature using K_2CO_3 as a base, but a reasonable yield (>60%) was realized using bis(2-iodoethoxy)ethane. Bis(2-iodoethoxy)ethane was generated *in situ* by treatment of **7** with excess lithium iodide at 80°C in *DMF* for 2 h. After cooling to ambient temperature, flame-dried K_2CO_3 and **6** were added (Scheme 1) and the reaction was stirred for 18 h. The product of the coupling was isolated from the reaction mixture by recrystallization of the solid residue obtained after concentrating the organic extracts. The ^1H NMR spectrum of **8** displayed the methylene units at $\delta = 4.54$ (t, $J = 5$ Hz), 4.31 (q, $J = 7$ Hz), 3.78 (t, $J = 5$ Hz) and 3.51 (s) ppm and the methyl groups at $\delta = 1.35$ (t, $J = 7$ Hz) ppm. Furthermore, its HRMS showed the molecular ion at $m/z = 482.1649$ (M^+) in agreement with the molecular formula $\text{C}_{20}\text{H}_{26}\text{N}_4\text{O}_{10}$.

Next, we proceeded to examine the reduction of the nitro groups in **8**. This was achieved employing Pd/C as a reduction system to furnish the corresponding unstable diamine that was coupled directly after concentration with 4-nitro-2-trichloroacetyl-1-methylpyrrole (**9**) [31] in the presence of triethylamine to afford the dinitro derivative **10** in 77% overall yield. This yield was obtained only when the solvents were degassed with N₂ prior to use. The product **10** was isolated by crystallization from ethanol. The structure was confirmed by spectral data and elemental analysis. The proton NMR spectrum displayed four signals with small coupling constant (2 Hz) representing the aromatic protons at $\delta = 8.18, 7.53, 7.44,$ and 6.91 ppm.

Securing **10** allowed further progress toward the projected final product. Thus, at this stage it was decided to introduce the dimethylaminopropyl group. This group is essential, because it plays an important role in the solubility of the molecule and its binding in the minor groove of the DNA. It was introduced by a two steps sequence. First, the ester groups were hydrolyzed to the corresponding dicarboxylic acid **11** in basic methanol. Second, the resulting dicarboxylic acid was further transformed into the corresponding diamide **12** by standard methodology implementing DCC in the presence of 3-hydroxy-1,2,3-benzotriazin-4(3*H*)-one (HOBt) and triethylamine.

Having completed the assembly of intermediate **12**, it remained to introduce the two pyrrole rings each having a formyl group at C-4. The methodology chosen for this task was designed to minimize the number of reaction steps required to transform **12** to the final product **5** due to the high polarity of this intermediate which caused difficulties in purification. This problem was solved by implementing the activated ester **13** [32]. Thus, the nitro groups in **12** were reduced to the corresponding unstable diamines using PtO₂ in methanol. This diamine was not isolated, but immediately allowed to react with excess activated ester **13** in DMF at 50°C in the presence of triethylamine to give the final product in reasonable yield (>65%). The target compound was purified on silica gel and eluted from the column using 5% NH₄OH in methanol:chloroform = 1:1. It was fully characterized using FABMS and NMR. Its FABMS showed the molecular ion at $m/z = 1079.8$ (MH⁺), in agreement with the molecular formula C₅₂H₇₀N₁₆O₁₀. The ¹H and ¹³C NMR spectrum of **5** provided conclusive evidence for the formation of the required target of the bisdistamycin analog **5**.

In conclusion, we have established a general synthetic route to a terminally linked bisdistamycin analog **5** starting from easily accessible starting materials. Biochemical analysis of interactions between **5** and DNA will be reported in due course.

Experimental

Melting points were determined on an Electrothermal melting point apparatus. Infrared (IR) spectra were recorded on a Nicolet 7199 FTIR spectrometer either in CHCl₃ or as KBr pellets. ¹H and ¹³C NMR spectra were recorded on Bruker AM-300 spectrometers using CDCl₃ and DMSO-d₆ as solvent and internal standard. Chemical shifts are reported in ppm relative to the residual solvent peak of CDCl₃ or DMSO-d₆, defined to be $\delta = 7.26$ and 2.49 ppm. For ¹³C NMR spectra (APT), (a) and (p) are used to denote the signals which are antiphase (methyl and methine groups) and in phase (methylene groups and quaternary carbons), respectively, relative to the solvent peaks. High-resolution mass

spectra (HRMS) and FAB mass spectra were obtained using a Kratos AEI MS-9 and MS-50 mass spectrometer. Elemental analyses were found to be in accord with the calculated values.

Bis[2-[(2-trichloroacetyl)-4-nitropyrrol-1-yl]ethoxy]ethane (8, C₂₀H₂₆N₄O₁₀)

A solution of 3.74 g bis(2-chloroethoxy)ethane (20.0 mmol) and 7.98 g LiI (60.0 mmol) in 150 cm³ DMF was stirred for 2 h at 80°C. After the solution was cooled to room temperature, 8.28 g K₂CO₃ (60.0 mmol) and 4.60 g of **6** (25.0 mmol) were added and the resulting mixture was stirred at room temperature for 18 h. The solvent was removed under reduced pressure at 50°C and the residual cake was washed with 3 × 300 cm³ CH₂Cl₂. The CH₂Cl₂ layer was washed with water, dried, and concentrated. The residue was recrystallized (20% *n*-hexane in CH₂Cl₂) to give 6.28 g (62%) pure **8**. Mp 102°C; ¹H NMR (CDCl₃): δ = 7.75 and 7.42 (2d, *J* = 2 Hz, 2 × 2pyrr-H), 4.54 (t, *J* = 5 Hz, 4H, 2NCH₂), 4.31 (q, *J* = 7 Hz, 4H, 2OCH₂), 3.78 (t, *J* = 5 Hz, 4H, 2CH₂), 3.51 (s, 4H, 2OCH₂), 1.35 (t, *J* = 7 Hz, 6H, 2CH₃) ppm; IR (CHCl₃ cast): $\bar{\nu}$ = 1715, 1539, 1508, 1317 cm⁻¹; HRMS: *m/z* = 482.1649 (M⁺).

Bis[2-[(2-carboethoxy)-4-[1-methyl-4-nitropyrrole-2-carboxamido]pyrrol-1-yl]ethoxy]ethane (10, C₃₂H₃₈N₈O₁₂)

A solution of 2.41 g **8** (5.0 mmol) and 30 mg 5% Pd/C in 300 cm³ 25% methanol in ethyl acetate was stirred under a hydrogen atmosphere (50 psi) until the starting material was consumed (18 h). After the catalyst was removed by filtration through celite, the filtrate was concentrated to dryness and the residual amine was dissolved in 50 cm³ DMF. Then 3.86 g **9** (15.0 mmol) and 3 cm³ triethylamine were added. The resultant mixture was stirred at 50°C for 3 h and then concentrated to dryness under reduced pressure at 50°C. The residue was recrystallized (ethanol) to give 2.82 g (77%) pure **10**. Mp 130°C; ¹H NMR (DMSO-*d*₆): δ = 10.21 (s, 2H, 2NH), 8.18, 7.53, 7.44, and 6.91 (4d, *J* = 2 Hz, 4 × 2pyrr-H), 4.40 (t, *J* = 5 Hz, 4H, 2NCH₂), 4.17 (q, *J* = 7 Hz, 4H, 2OCH₂), 3.92 (s, 6H, 2NCH₃), 3.63 (t, *J* = 5 Hz, 4H, 2OCH₂), 3.50 (s, 4H, 2OCH₂), 1.25 (t, *J* = 7 Hz, 6H, 2CH₃) ppm; IR (CHCl₃ cast): $\bar{\nu}$ = 1689, 1660, 1565, 1379 cm⁻¹; HRMS: *m/z* = 726.2631 (M⁺).

Bis[2-[(2-carboxy)-4-[1-methyl-4-nitropyrrole-2-carboxamido]pyrrol-1-yl]ethoxy]ethane (11, C₂₈H₃₀N₈O₁₂)

20 cm³ 2 *N* NaOH was added to a solution of 3.63 g **10** (5.0 mmol) in 50 cm³ methanol and the resultant mixture was stirred at 50°C for 4 h. The reaction mixture was cooled to 0°C and the *pH* was adjusted to 2 with conc HCl. The precipitate was collected and dried to furnish 2.4 g (71%) pure **11**. Mp 157°C; ¹H NMR (DMSO-*d*₆): δ = 12.50 (s, 2H, 2COOH), 10.20 (s, 2H, 2NH), 8.15, 7.50, 7.45, and 6.84 (4d, *J* = 2 Hz, 4 × 2pyrr-H), 4.43 (t, *J* = 5 Hz, 4H, 2NCH₂), 3.93 (s, 6H, 2NCH₃), 3.64 (t, *J* = 5 Hz, 4H, 2OCH₂), 3.48 (s, 4H, 2OCH₂) ppm; IR (KBr): $\bar{\nu}$ = 1658, 1565, 1455 cm⁻¹; FABMS: *m/z* = 670.8 (M⁺).

Bis[2-[[2-(carboxamidopropyl)-3-dimethylamino]-4-[1-methyl-4-nitropyrrole-2-carboxamido]pyrrol-yl]ethoxy]ethane (12, C₃₈H₅₅N₁₂O₁₀)

A solution of 1.67 g **11** (2.5 mmol), 1.43 g EDCI (7.5 mmol), and 1.22 g 3-hydroxy-1,2,3-benzotriazin-4(3*H*)-one (HOBt) (7.5 mmol) in 50 cm³ DMF was stirred at room temperature for 1 h. Then 3 cm³ 3-dimethylaminopropylamine and 3 cm³ triethylamine were added and the resulting mixture was stirred for 3 h. The solvent was removed under reduce pressure at 50°C and the residue was dissolved in 300 cm³ CHCl₃. The solution was washed with 20 cm³ 5% K₂CO₃ and the organic layer was separated, dried, and concentrated. The crude product was purified on silica gel (flash chromatography, 5% NH₄OH in methanol:CHCl₃ = 1:1) to furnish 1.10 g (52%) **12**. Mp 179°C; ¹H NMR (CDCl₃): δ = 8.90 (s, 2H, 2NH), 7.95 (bs, 2H, 2NH), 7.55, 7.34, 7.18, and 6.75 (4d, *J* = 2 Hz, 4 × 2pyrr-H), 4.43 (t, *J* = 5 Hz, 4H, 2NCH₂), 4.02 (s, 6H, 2NCH₃), 3.69 (t, *J* = 5 Hz, 4H, 2OCH₂), 3.49 (s, 4H, 2OCH₂), 3.45 (q, *J* = 6 Hz, 4H, 2NCH₂), 2.50 (t, *J* = 6 Hz, 4H, 2NCH₂), 2.32 (s, 12H, 2N(CH₃)₂), 1.73 (m, 4H, 2CH₂) ppm; IR (CHCl₃ cast): $\bar{\nu}$ = 1639, 1592, 1573, 1504 cm⁻¹; FABMS: *m/z* = 839.1 (MH⁺).

Bis[2-[2-(carboxamidopropyl-3-dimethylamino)[1-methyl-4-[1-methyl-4-formamidopyrrole-2-carboxamido]pyrrole-2-carboxamido]pyrrol-yl]ethoxy]ethane (5, C₅₂H₇₁N₁₆O₁₀)

A solution of 209 mg **12** (0.25 mmol) and 30 mg PtO₂ in 100 cm³ methanol was stirred under hydrogen (50 psi) for 3 h. Then, the catalyst was removed by filtration through celite. The filtrate was concentrated and the residue was dissolved in CHCl₃. The residue left after concentration was dissolved in 20 cm³ DMF. The resulting solution was added to 314 mg **13** (0.10 mmol) followed by 1 cm³ triethylamine. The resulting mixture was stirred for 6 h and then the solvent was removed under reduced pressure at 50°C. The residue was purified on silica gel (flash chromatography, 5% NH₄OH in methanol:CHCl₃ = 1:1) to give 185 mg (68%) pure **5**. Mp 184°C; ¹H NMR (DMSO-d₆): δ = 10.05, 9.93, and 9.90 (3 s, 3 × 2NH), 8.15 (d, *J* = 2 Hz, 2H, 2HCO), 8.10 (t, *J* = 5 Hz, 2H, 2NH), 7.24, 7.23, 7.18, 7.03, 6.92, and 6.85 (6d, *J* = 2 Hz, 6 × 2pyrr-H), 4.40 (t, *J* = 5 Hz, 4H, 2NCH₂), 3.84 (s, 12H, 4NCH₃), 3.61 (t, *J* = 5 Hz, 4H, 2NCH₂), 3.45 (s, 4H, 2OCH₂), 3.18 (q, *J* = 7 Hz, 4H, 2CH₂), 2.24 (t, *J* = 7 Hz, 4H, 2NCH₂), 2.13 (s, 12H, 2N(CH₃)₂), 1.60 (quin, *J* = 7 Hz, 4H, 2CH₂) ppm; ¹³C NMR (DMSO-d₆): δ = 161.2 (p), 158.4 (p), 158.3 (p), 157.8 (a), 122.9 (p), 122.8 (p), 122.6 (p), 122.5 (p), 122.2 (p), 122.0 (p), 120.7 (a), 118.4 (a), 117.4 (a), 104.6 (a), 104.5 (a), 103.9 (a), 70.7 (p), 69.5 (p), 56.9 (p), 47.3 (p), 45.0 (a), 37.0 (p), 36.3 (a), 36.0 (a), 27.0 (p) ppm; IR (CHCl₃ cast): $\bar{\nu}$ = 3285, 1641, 1580, 1527, 1435 cm⁻¹; FABMS: *m/z* = 1079.8 (MH⁺).

Acknowledgements

We thank the Deanship of Research at Jordan University of Science and Technology for financial support.

References

- [1] Arcamone F, Orezzi PG, Barbier W, Nicolella V, Penco S (1967) *Gazz Chim Ital* **97**: 1097
- [2] Arcamone F, Penco S, Orezzi PG, Nicolella V, Pirelli A (1964) *Nature* **203**: 1064
- [3] Arcamone F, DiMarco A, Scott T (1961) *Microbio* **9**: 83
- [4] Zimmer C (1975) *Prog Nucleic Acid Res Mol Biol* **15**: 285
- [5] Lown JW (1992) *Antiviral Research* **17**: 179
- [6] Kopka ML, Yoon C, Goodsell DS, Pjura P, Dickerson RE (1985) *Proc Nat Acad Sci USA* **82**: 1376
- [7] Rao KE, Shea RG, Yadagiri B, Lown JW (1990) *Anticancer Drug Des* **5**: 3
- [8] Kopka ML, Goodsell DS, Han GW, Chiu TK, Lown JW, Dickerson RE (1997) *Structure* **5**: 1033
- [9] Nguyen DH, Szewczyk JW, Baird EE, Dervan PB (2001) *Bioorg Med Chem* **9**: 7
- [10] Sharma SK, Tandeon M, Lown JW (200) *J Org Chem* **65**: 1102
- [11] Zhao R, Al-Said NH, Sternbach DL, Lown JW (1997) *J Med Chem* **4**: 216
- [12] Baradli PG, Balboni G, Cacciari B, Guitto A, Manfredini S, Romagnoli R, Spalluto G, Thurston DE, Howard PW, Bianchi CR, Mischiati C, Gambari R (1999) *J Med Chem* **42**: 5131
- [13] Damayanthi Y, Praveen BS, Lown JW (1999) *J Org chem* **64**: 290
- [14] Cozzi P (2000) *Il Farmaco* **55**: 168
- [15] Baradli PG, Nunez MC, Espinosa A, Romagnoli R (2004) *Current Topics in Med Chem* **4**: 219
- [16] Goodsell D, Dickerson RE (1986) *J Med Chem* **29**: 727
- [17] Mrksich M, Dervan PB (1993) *J Am Chem Soc* **115**: 4472
- [18] Geierstanger B, Jacobson JP, Mrksich M, Dervan PB, Wemmer DE (1994) *Biochemistry* **33**: 3055
- [19] Pelton JP, Wemmer DE (1990) *J Am Chem Soc* **112**: 1393
- [20] Pelton JP, Wemmer DE (1989) *Proc Nat Acad Sci USA* **86**: 5723
- [21] Wade WS, Mrksich M, Dervan PB (1992) *J Am Chem Soc* **114**: 8783

- [22] Wemmer DE (2000) *Annu Rev Biophys Biomol Struct Sc* **29**: 439
- [23] Wang W, Lown JW (1992) *Med Chem* **35**: 2890
- [24] Lown JW, Krowicki K, Newman RA, De Clereq E (1989) *J Med Chem* **32**: 2368
- [25] Baligna R, Baired EE, Herman DM, Melander C, Dervan PB, Crothers DM (2001) *Biochemistry* **40**: 3
- [26] Al-Said NH, Lown JW (1994) *Tetrahedron Lett* **35**: 7577
- [27] Al-Said NH, Lown JW (1995) *Synthetic Commun* **25**: 1095
- [28] Chen YH, Yang Y, Lown JW (1996) *J Biomolec Struct Dyn* **14**: 341
- [29] Sharma SK, Tandeon M, Lown JW (2000) *J Org Chem* **65**: 1102
- [30] Sharma SK, Tandeon M, Lown JW (2002) *Tetrahedron* **58**: 3417
- [31] Belanger P (1979) *Tetrahedron Lett* **27**: 2505
- [32] Grehn L, Ragnarson U (1981) *J Org Chem* **46**: 3492